

The Comparative Study of Antioxidants in Plants of East Georgian Arid Habitats (Yagljudja and Kvernaqi hills)

Gulnara Badridze*

Department of Botany, Ilia State University, Botanikuri, Republic of Georgia

Research Article

Received: 10-May-2022,

Manuscript No. JEAES-22-63012;

Editor assigned: 14- May-2022, Pre

QC No. JEAES-22-63012 (PQ);

Reviewed: 28- May-2022, QC No.

JEAES-22-63012; **Revised:** 07-Jun-

2022, Manuscript No. JEAES-22-

63012 (A); **Published:** 14-Jun-2022,

DOI:10.4172/23477830.10.04.001

***For Correspondence:** Gulnara Badridze, Department of Botany, Ilia State University, Botanikuri,

Republic of Georgia; Tel: +995

555270698

E-mail: gbadridze@yahoo.com;

Keywords: Drought resistance; Antioxidants; Preliminary report

ABSTRACT

The preliminary report of 2021 of the world meteorological organization on the climate global situation clears its negative changes all over the world. Especially disturbing is the temperature rise, accompanied by hot waves, forest fires, intensive melting of the ice cover, and other undesirable aftereffects. Climate warming significantly increases the chances of dying of many plant species. Drought resistant plants have the highest potential of adaptation to increased temperature and water deficiency. Thus, knowledge of their biology will be especially important in deserted regions restoration. The nonspecific mechanisms of resistance, especially antioxidant system, are concerned as one of the leading in plants drought resistance. The presented study aimed the comparative study of the indices of antioxidant system (ascorbic acid, tocopherol, carotenoids, anthocyanins, soluble phenols, proline, total proteins and soluble carbohydrates, as well as the activity of catalase, peroxidase, nitrate reductase and the total antioxidant activity in percents of inhibition) of drought resistant species-*Astragalus microcephalus* Willd. (*Astracantha microcephala* (Willd.) Podlech)-goat's thorn, *Theucrium polium* L-felty germander, *Euphorbia seguieriana* Neck-spurge, *Capparis spinosa* L-caper bush, *Paliurus spina-christi* Mill-Christ's thorn, growing in two different arid habitats of the East Georgia (Yagljudja and Kvernaqi hills).

The defence mechanisms of the antioxidant system appeared to be partially different in one and the same species of various habitats, as well as in different species of the same habitats. Activation of phenolic substances and anthocyanins synthesis against extreme conditions of both habitats (water deficiency, high temperature and intensive irradiation) was common for all tested species. Additionally, activation of peroxidase in Kvernaqi species and intensive accumulation of soluble carbohydrates in Iagljudja plants was mentioned.

INTRODUCTION

The 26th conference of the UNO frame convention on climate change (COP26) held in Glasgow in October-November of 2021 is one additional warning to mankind about undesirable results of the climate global change. The preliminary report of the world meteorological organization on the climate global situation in 2021 presented the latest scientific documentations demonstrating evident negative changes all over the world. Especially disturbing is the temperature rise, accompanied by hot waves, forest fires, intensive melting of the ice cover, and other undesirable aftereffects. Last seven years were regarded as the hottest through the whole history of climate observation [1]. Climate warming significantly raises the risk of plants dying off under the increased stresses [2,3]. Presumably the area of distribution of many plant species will change. The migration rate will depend on species features, competition, climate conditions, etc [4]. Drought resistant species will have the highest potential of adaptation to increased temperatures and accompanying water deficiency. Thus, the knowledge of their biology will be very important in the restoration of deserted areas; moreover, most of them are used in medicine. Drought resistant plants possess evolutionary developed physiological and biochemical mechanisms of stability against water deficiency and high temperature. The nonspecific mechanisms of resistance are regarded as one of the principle under stress conditions; the antioxidant system is of special importance among them [5,6]. Characteristics of the antioxidant system of drought resistant plants of the arid territories of Georgia are practically unexplored. The presented work aimed comparative studying of some characteristics of drought resistant species growing at different arid habitats of east Georgia (Iagljudja and Kvernaqi hills). The study may be regarded as the continuation of previous year's investigations [7]. Content of ascorbic acid, tocopherol, carotenoids, anthocyanins, soluble phenols, proline, total proteins and soluble carbohydrates, as well as the activity of catalase, peroxidase, nitrate reductase and the total antioxidant activity in percents of inhibition have been studied in leaves of experimental plants.

MATERIALS AND METHODS

Research area

Experimental species were collected in July of 2020-21, at two different arid habitats of East Georgia Iagljudja hill (Gardabani municipality) and Kvernaqi hill (Kaspi municipality). Iagljudja hill is situated in Kvemo Kartli (lower Kartli), on the north-east of Marneuli plane, near the city Rustavi. The climate here is continental, with mild winter and hot, dry summer. The mean annual temperature is 12°C-13°C; in the coldest month January the mean temperature is 0.3°C-0°C. Especially hot is July and August. Absolute minimal temperature is minus 20°C-25°C and maximal 40°C-41°C. The mean annual amount of precipitations is 350-500 mm [8]. Most part of Marneuli municipality soils are degraded to various degrees; this is clear from the worsening of their physical and mechanical, chemical and

microbiological properties and fertility decrease. Degradation of a plant cover resulted in a formation of clay-rich, low-humic, calcareous grey-brown soils (Official web-page of Marneuli municipality). Kvernaqi hill is situated in Shida Kartli (inner Kartli), near the city Kaspi. Climate here is transitional from subtropical to humid. The winter is moderately cold, summer is dry and hot. The mean annual temperature of air is 11.4°C, in January minus 0.5°C, in August 23°C. The absolute minimal temperature is minus 27°C, and absolute maximal 40°C. Annual amount of precipitations makes 450 mm. On the south slope of Kvernaqi hill (where the material was collected) the radiation is intensive in summer; that is why the air temperature here is higher compared to the northern slope. In spite of comparatively high precipitations in Kvernaqi, compared to central regions of Shida Kartli plane, their efficiency here is low; because of it the soil is dryer. Kvernaqi hill is constructed of Neogenic conglomerates, sandy-gravel shales. In Kotsakhura gorge, where the material was picked, soils are alluvial, here and there rocky and gravel [8,9].

Experimental plants

Middle age, mature, healthy leaves were collected at least from 5 different individuals of each experimental species: *Astragalus microcephalus* Willd. (*Astracantha microcephala* (Wild) Podlech)-goat's thorn, *Theucrium polium* L.-felty germander, *Euphorbia seguieriana* Neck. Spurge, *Capparis spinosa* L. Caper bush, *Paliurus spina* Mill.-Christ's thorn. Material was taken at 530-395 m above sea level; in fruit bearing phase, during the hottest period for these locations (38°C-40°C). Analyses were performed both on raw and dry material, with 3-fold repetitions.

***Astragalus microcephalus* wild:** (*Astracantha microcephala* (Willd) (Podlech) goat's thorn (*Fabaceae*) is representative of one of the largest genus. Plants of the genus are widely used in medicine, as food and fodder, fuel, for ornamental purposes, etc. Most species, among them *A. microcephalus*, are spread in arid and semi-arid habitats of the world. *A. microcephalus* is one of the popular species of the genus, and is used for extracting tragacanth gum [10,11].

***Theocrium polium* L:** Felty germander (*Lamiaceae*) is a perennial herb, spread in rocky and sandy, semi-arid places of mediterranean region, Europe, North Africa and south-east Asia; it is widely used both in traditional and folk medicine [11-14].

***Euphorbia seguieriana* neck:** Spurge (*Euphorbiaceae*) is a perennial herb; it grows on sandy and stony slopes, limestone soils, river dry beds, and semi-desert and steppe zone. The plant is xeromezophyte; grows in many parts of the world [15]. The genus *Euphorbia* comprises plants of high economic value with different morphology, and inhabiting various environmental conditions. Species of the genus, among them *E. seguieriana* have multilateral medicinal use. The phytochemistry of the genus is still intensively studied [16].

***Capparis spinosa* L:** Caper bush (*Capparidaceae*) is a xerophyte plant with deep root system, which supports it to stand long period drought and high temperatures. The plant grows in clay and sandy deserts, gravel-stony places. It is recommended as one of the species for restoration of saline and eroded soils. *C. spinosa* is popular as food and medicinal plant [17,18].

***Paliurus spina* mill:** Christ's thorn (*Rhamnaceae*) is a perennial, deciduous thorny bush, widely spread in Mediterranean region, and at dry and rocky places of Asia. The plant is less demanding to soil and grows even on an

alkali one. Christ's thorn is drought resistant and inhabits degraded areas; it supports the soil and protects it against erosion. Different parts of the plant are used both in traditional and folk medicine [19-21].

Antioxidant enzymes assay

Peroxidase activity was determined spectrophotometrically: optical density of the products of guaiacol oxidation was measured at the wave length of 470 nm by the spectrophotometer (SPEKOL 11, KARL ZEISS, and Germany) [22]. Catalase activity was studied gasometrically: volume of the oxygen released in the process of reaction between hydrogen peroxide and enzyme was measured [23].

Nitrate reductase assay: Method of determining the nitrate reductase activity was based on measurement of nitrites amount, which were formed as a result of nitrate reductase reaction with the infiltrated nitrates [22].

Ascorbic acid

A titration method was used to measure the content of ascorbic acid in plant material. 2 g of fresh leaves were mashed in 15 ml of 2% hydrochloric acid and 10 ml of 2% metaphosphoric acid, and filtered. One ml of the filtrate was added to 25 ml of distilled water and titrated with a 0.001 M solution of dichlorophenolindophenole [22].

Tocopherol

Two grams of ground leaves were extracted with 20-25 ml of pure ethanol (three-fold). The combined extract was mixed with 20 ml of 60% potassium hydroxide, and saponificated on water bath for 2hrs. Tocopherol was extracted from the obtained hydrolyzate using diethyl-ether (3-fold extraction). The combined extract was washed with distilled water until a complete removal of alkaline residuals. Water was removed with Na₂SO₄; the obtained solution was evaporated on the water bath, cooled, mixed with alcohol-nitric acid (1 ml of concentrated HNO₃:5ml of 96° alcohol), and boiled during 3 min till the color became dark red. Extinction of the extract was measured at 470nm by the spectrophotometer (SPEKOL 11, KARL ZEISS, and Germany) [24].

Anthocyanins

100 mg of grinded leaves were added with 20 ml of 96% acidified (with 1% HCl) ethanol (99:1). After 24hrs retention in dark the optical density at 540 nm was measured (spectrophotometer SPEKOL 11, KARL ZEISS, Germany) [22].

Plastid pigments

Chlorophylls and carotenoids were determined spectrophotometrically. Fresh leaves (100-200 mg) were mashed with sand and CaCO₃ and washed with ethanol. Optical density of the filtrate was measured (spectrophotometer SPEKOL 11, KARL ZEISS, Germany). Concentration of chlorophylls a and b, also carotenoides was calculated by the formula of Wintermanns [25].

Total phenols

A 0.5 g of fresh leaves was boiled in 80% ethanol for 15 min. After centrifugation the supernatant was saved, and residues of leaves were mashed in 60% ethanol and boiled for 10 min. Obtained extract was added to the first supernatant and evaporated. The sediment was dissolved in distilled water. One ml of the received solution was added with the Folin-Ciocalteu reagent and optical density was measured at 765 nm. The chlorogenic acid served as control [26].

Total protein assay

Content of proteins was determined after Lowry (1951).

Proline: 0.5 g of dry leaves were mashed in 10 ml of 3% sulphosalicylic acid and filtered. 2 ml of the filtrate was added to 2 ml of acid ninhydrin and 2 ml of ice acetic acid. After 1 hr exposition on a water bath the extract was cooled and added with 4 ml of toluene and divided in a separating funnel. Optical density of upper layer was measured on a spectrophotometer (SPEKOL 11, KARL ZEISS, and Germany) at 520 nm ^[27].

Soluble carbohydrates: Content of soluble carbohydrates was tested with anthrone reagent ^[28]. To 100 mg of air-dry leaf material was added 96° alcohol for extraction (3-fold). The total amount of the obtained extract was evaporated on a water bath and dissolved in 5 ml of distilled water. To 0.5 ml of the tested water extract was added 2 ml of anthrone reagent and heated in a water bath for 10 min. After this procedure the test-tubes were placed in a cold water bath and 15 min later the optical density of the solution was measured at 620 nm with a spectrophotometer (SPECOL 11, KARL ZEISS, and Germany).

Nitrates: After the water-extraction of 500 g of plant material (homogenized for 30 min at room temperature), it was filtered. Hydrogene peroxide was added to 10 ml of the filtrate and evaporated. Disulphophenolic acid was added to the obtained sediment and optical density was determined at 410 nm (SPEKOL 11, KARL ZEISS, Germany) ^[23,29].

Total antioxidant activity: This index was measured by modified method using Diphenyl-Picryl-Hydrazyl (DPPH) ^[30]. 200 mg of experimental powder was extracted with 96° ethanol (two-fold). The obtained extract was evaporated on a water bath and the sediment was dissolved in 10 ml of water-alcohol mixture. The 0.01 ml of the received solution was added with 4 ml of 40 µM DPPH solutions and after 30 minutes of incubation in the dark, the optical density was measured at 515 nm by the spectrophotometer (SPEKOL 11, KARL ZEISS, and Germany). The percent of inhibition was calculated.

Statistical processing of data: One way ANOVA and Tukey's multiple comparison tests were used to test differences between the means. All calculations were performed using statistical software Sigma Plot 14.5.

RESULTS AND DISCUSSION

As it was mentioned above, the experimental plants were collected in July, which is the hottest and driest month of the studied habitats. Also the fact that the last seven years are regarded as the hottest through the whole history of climate observation must be taken into account ^[1]. Thus, the tested species had to survive under combined stress of high temperature, water deficiency and intensive irradiation simluteniously.

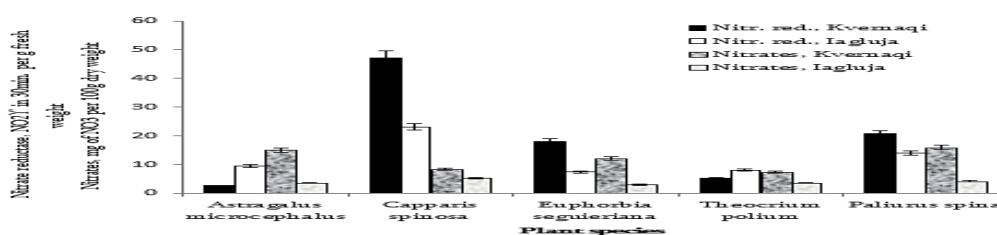
Nitrate reductase and nitrates

Nitrate reductase (EC 1.6.6.1.) plays an essential role in plants physiological and metabolic condition as it is a key enzyme in synthesis of amino acids, proteins, chlorophylls and other nitrogen-containing substances; it may serve as a marker of drought stress ^[31-33]. It is established that the activity of nitrate reductase decreases under the water deficiency; which is associated with photosynthesis inhibition ^[34]. Moreover, as the nitrate reductase is the substrate-dependent enzyme, and its activity is regulated by the concentration of nitrates and ammonium, the decline of enzyme's activity may be caused by reduction of nitrates in leaves ^[35,36]. As the absorption of water and

minerals, among them of nitrates, is reduced during the drought, this will be reflected on enzyme's activity as well [34,37]. According to the experimental results it is clear that the one and the same species of two habitats showed statistically different activity of nitrate reductase ($p \leq 0.002$); indices of goat's thorn and feltly germander growing at lagludja were respectively 3.6 and 1.6 times higher, compared to Kvernaqi results ($p \leq 0.002$). In other tested species on the contrary Kvernaqi data of nitrate reductase activity were higher: In caper bush 2 times, in spurge 2.5 times, and in Christ's thorn 1.5 times. Although the activity of nitrate reductase has been associated with nitrates content in leaves [35,36], in our experimental plants this correlation was not clear. Generally, the content of nitrates in Kvernaqi species was higher that of lagludja ones ($p < 0.05$) (Figure 1). Moreover, in lagludja experimental species content of nitrates was low and statistically similar, while the same species of Kvernaqi statistically differed by this index ($p < 0.05$). If we consider the content of chlorophylls as the indicator of photosynthetic activity, there may be assumed some relationship between nitrate reductase activity and chlorophylls content in leaves; as the decrease of the enzyme's activity is associated with photosynthesis inhibition under the water deficiency [34]. Comparision of nitrate reductase activity and chlorophylls content in tested plants revealed the coincidence of the decrease of these two indices (Figures 1 and 2). As the nitrate reductase gives some idea on the general physiological condition of a plant, according to the obtained results it may be concluded that lagludja conditions were more stressful for experimental plants, than Kvernaqi. Equally low data of nitrates in lagludja species is demonstration of this (Figure 1).

Figure 1. Nitrate reductase activity and content of nitrates in leaves of Kvernaqi and lagludja (East Georgia) plants.

Note: (■) Nitr. red, kvemaqi; (□) Nitr.red, lagludja; (▨) Nitrates, Kvernaqi; (□) Nitrates, lagludja

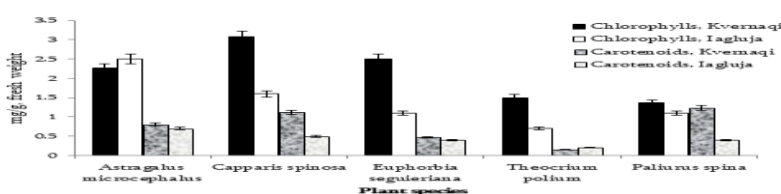


Chlorophylls and carotinoids

It is known that water deficiency, together with intensive radiation and high temperature inhibits photosynthesis. One of the reasons is chloroplasts damage and chlorophylls decrease, regarded as some kind of protective reaction towards stresses [37-40]. Comparative study of chlorophylls and carotenoids in one and the same species of the studied habitats has revealed the statistical similarity of chlorophylls in leaves of goat's thorn feltly germander and Christ's thorn ($p=0.4$, $p=0.2$ and $p=0.7$ respectively), while in other species it was different ($p < 0.05$). Moreover, Kvernaqi data were higher than lagludja ones: in caper bush 1, 9 times, in spurge 2, 3 times (Figure 2). The between-species comparison of results revealed statistical similarity of chlorophylls in spurge and goat's thorn ($p=0.6$), caper bush and spurge ($p=0.1$), feltly germander and Christ's thorn ($p=0.5$) growing at Kvernaqi hill; while among lagludja species chlorophylls content of leaves was statistically similar in all tested species ($p < 0.05$), except

goat's thorn. It must be mentioned that inspite of stressful conditions in leaves of both habitat plants content of chlorophylls was not low; which indicates to reliable protection of the photosynthetic apparatus. As for carotenoids, between-habitat differences were revealed in caper bush ($p=0.03$) and Christ's thorn ($p=0.06$) individuals. Between-species comparison cleared statistical similarity of carotenoids in caper bush and Christ's thorn leaves ($p=0.4$) growing at Kvernaqi; while at lagludja statistically same were indices of goat's thorn and caper bush ($p=0.06$), as well as of all other species ($p>0.05$) (Figure 2).

Figure 2. Content of chlorophylls and carotenoids in leaves of Kvernaqi and lagluja (East Georgia) plants. Note: (■)Chlorophylls, Kvernaqi;(□) Chlorophylls,lagluja; (▨)Carotenoids, Kvernaqi;(□) Carotenoids, lagluja

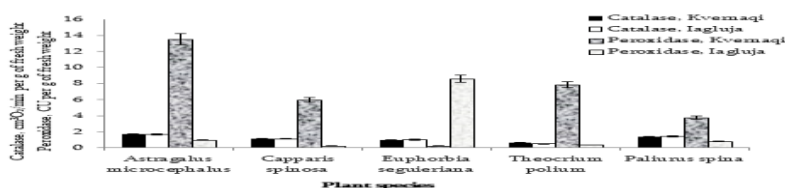


Catalase and peroxidase

Generally the high activity of antioxidant enzymes-catalases and peroxidases has been mentioned in drought resistant plants [6,34]. Although the high specificity of the activity is characteristic for enzymatic antioxidants-they affect particular type of radicals, with specific cellular and organ localization [41]. Catalases of the photosynthesizing tissue mainly are localized in peroxisomes (CAT, EC 1.11.1.6), where they destruct the hydrohen peroxide produced during photorespiration [42]; Moreover, proliferation of peroxisomes and diffusion additional hydrogen peroxide from the cytosol may take place under stress, which is actively neutralized by catalases as well [43]; Although, in some plants decrease of catalases activity is possible, which is regared as a demonstration of a significant role of peroxidases and ascorbate-glutathione cycle, as of principal oxygen-scavengers [44]. So called classic, or non-specific peroxidases (EC 1.11.1.7) are multifunctional enzymes, which like catalases, neutralize the active forms of oxygen. For this purpose they use different reducing agents, often phenolic substances. They are concentrated mainly in a vacuole and cytosol [45]. In studied species the activity of catalase was generally low. Comparison of results of one and the same species of two habitats revealed their statistical identity ($p>0.05$). Among Kvernaqi plants statistically similar activity of catalase was found in goat's thorn and Christ's thorn ($p=0.09$), caper bush and Christ's thorn ($p=0.4$), caper bush and spurge ($p=0.2$). Among laglidja species statistically similar activity of catalase was revealed in caper bush, spurge and Christ's thorn ($p=0.5$). The peroxidase activity of same species from different habitats was statistically different ($P \leq 0,001$). Generally the peroxidase activity of Kvernaqi experimental species was significantly higher, compared to lagludja data (except spurge): in goat's thorn leaves it was 14 times higher, in caper bush 26 times, in felty germander 22 times, and in Christ's thorn -4.5-

times higher. While comparing peroxidase activity of different species, among lagludja plants spurge and Christ's thorn results were similar ($p=0.5$), in other species there was statistical difference ($p<0.05$). In Kvernaqi plants all data on peroxidase activity were statistically diverse ($p<0.01$) (Figure 3).

Figure 3. Activity of catalase and peroxidase in leaves of Kvernaqi and lagluja (East Georgia) plants. Note: (■) Catalase, Kvernaqi; (□) Catalase, lagluja; (▨) Peroxidase, Kvernaqi; (▩) Peroxidase, lagluja



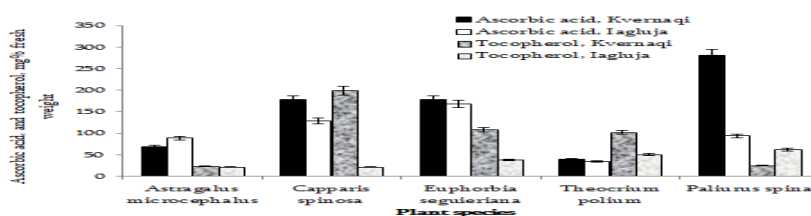
Analyzing the experimental results it may be supposed that low catalase activity in plants of both habitats indicates to the low concentration of hydrogen peroxide in leaves. Since catalase has a low affinity to hydrogen peroxide, compared to peroxidase, it is active under high concentrations of the substrate [46,47]. Decreased activity of catalase may be caused by its inactivation because of intensive radiation as well [41]. High activity of peroxidase in experimental plants of Kvernaqi hill may be explained if we take into account the fact that peroxidases are active in those systems, where the primary transformation of derivatives of hydrohen peroxide organic peroxides takes place by means of appropriate reducing agents [46]. High peroxidase activity of spurge individuals at lagludja, as exception, may be linked with the diverse from other species C_4 way of CO_2 assimilation in this species. Thus, more extreme conditions of lagludja hill compared to Kvernaqi, cause diverse biochemical changes in experimental species. It may be supposed that the enzymatic antioxidant system of lagludja experimental plants is not leading in adaptation to the intensive irradiation, high temperature and water deficiency conditions of this habitat; while under Kvernaqi conditions peroxidase was distinguished from this point of view.

Ascorbic acid and tocopherol

Ascorbic acid is regarded as one of the leading substances for protection of the photosynthetic apparatus against the oxidative stress [48]. Under the intensive irradiation as well as drought, content of ascorbic acid in plant increases. High content of the substance is one of the features of plant stress-resistance [49,50]. In a number of plants the increase of tocopherol, together with ascorbic acid, has been indicated as one of the primary reactions against drought or intensive radiation stress [51,52]. Among one and the same species of different habitats the content of ascorbic acid was statistically similar in leaves of spurge and felty germander ($p=0.1$ and $p=0.8$ respectively), while in other species it was different ($p<0.05$); in particular, Kvernaqi indices were higher compared

to lagludja: in caper bush by 28%, in spurge by 6%, in Christ's thorn by 66.5%. In leaves of goat's thorn in contrary it was by 22.5% lower (Figure 4). According to obtained results it may be supposed that in leaves of caper bush, spurge and Christ's thorn, growing in Kvernaqi conditions, ascorbate system is one of the leading mechanisms in leaves protection against stress; while under lagludja conditions this mechanism was clearly expressed only in caper bush and spurge leaves. It must be mentioned that chloroplast membrane contains high amount of α -tocopherol, which protects it against photooxidation. By well-known ascorbate-tocopherol cycle the oxidized tocopherol is reduced by means ascorbic acid [53,54]. Among the same species of studied habitats content of tocopherol was statistical identical only in goat's thorn leaves ($p=0.109$); in other experimental species lagludja data were higher compared to Kvernaqi ones ($p<0.05$): in caper bush 9 times, in spurge 2, 8 times, in felty germander 2 times; in Christ's thorn-incontrary Kvernaqi index was 2,5 times higher; According to results it may be supposed that increase of the tocopherol synthesis in leaves of caper bush, spurge and felty germander is one of the stress-protective mechanisms under lagludja conditions, while in Kvernaqi this mechanism was essential only in Christ's thorn (Figure 4).

Figure 4. Content of ascorbic acid and tocopherol in leaves of Kvernaqi and lagludja (East Georgia) plants. Note: (■)Ascorbic, Kvernaqi;(□) Ascorbic acid, lagludja; (▨) Tocopherol, Kvernaqi;(□) Tocopherol, lagludja

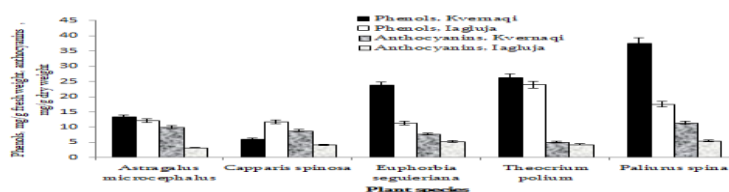


Soluble phenols and anthocyanins

Among the low-molecular antioxidants phenolic substances are distinguished by the active interaction with free radicals. Accordingly, their role in the defence of cell membrane against the oxidative stress is very high [55,56]. Some papers deal with the phenolics accumulation in plant cell under water deficiency [57]. It must be mentioned that generally high content of phenols was revealed in plants of both habitats; although in same species of the two habitats statistical similar results were received only in goat's thorn and felty germander ($p=0.08$) (Figure 5). As for other species, Kvernaqi results of spurge and Christ's thorn were 2 times higher compared to lagludja data ($p<0.05$), while in caper bush in contrary lagludja results were 2 times higher. According to results it is clear that high content of phenolics in both habitats is one of the leading stress-protective mechanisms in tested species. One of the groups of phenolic substances anthocyanins, which are partially concentrated in leaf epidermis, plays a role of light reflector, protecting chlorophyll from excess radiation [58]. It has been established that accumulation of anthocyanins in vegetative tissues increases under various stresses (drought, intensive radiation, etc.) [58-60]. It is supposed that they take part in diminuation of cell osmotic potential and thus retain water and turgor in the cell [61].

High content of anthocyanins was mentioned in experimental plants of both studied habitats. Although, while comparing data of one and the same species growing at different locations, it is evident that in Kvernaqi individuals anthocyanins were higher, compared to lagludja ($p \leq 0.001$); in particular results of goat's thorn were 3 times higher, that of caper bush and Christ's thorn 2 times, and of spurge 1, 5 times higher. According to experimental results it may be assumed that accumulation of anthocyanins, especially in Kvernaqi plants demonstrates the leading stress-protective role of these substances (Figure 5).

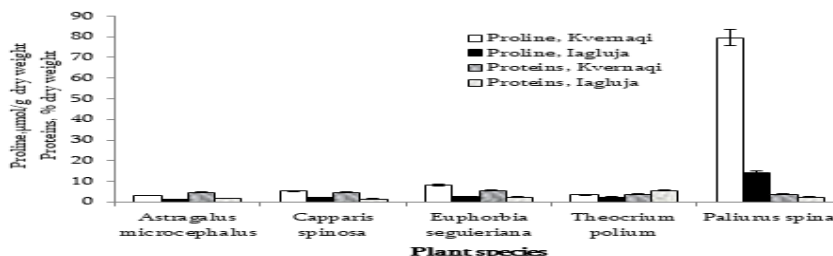
Figure 5. Content of polyphenols and anthocyanins in leaves of Kvernaqi and lagludja (East Georgia) plants. Note: (■) Phenols, Kvernaqi; (□) Phenols, lagludja; (▨) Anthocyanins, Kvernaqi; (□) Anthocyanins, lagludja



Proline, total proteins and soluble carbohydrates

Other metabolites may also take part in plant protection against oxidative stress, together with antioxidants. These substances (amino acids, soluble carbohydrates, proteins) so called osmoprotectants, are not antioxidants, but reveal similar properties and reliably protect lipo-protein components of membranes against damage [62-64]. Many authors point to a positive role of amino acid proline in drought-resistance of plant [65,66]. It holds water in the cell and retains its turgor [67,68]. Individuals of one and the same species, growing at different habitats had statistically different content of proline in leaves ($p \leq 0.01$); moreover, in lagludja species the content of amino acid was lower compared to Kvernaqi: in goat's thorn 2, 7 times lower, in caper bush 2, 5 times, in spurge 3, 3 times, and in Christ's thorn 5,6 times lower. Only data of felted germander coincided in both habitats ($p=0.3$) (Figure 6). Generally the content of proline in plants of both habitats was not high. Exception was Christ's thorn; especially Kvernaqi individuals of this plant were distinguished with high content of proline in leaves. Thus, it may be concluded that the stress-protective function of proline was not principal in tested plants, except Christ's thorn. Adaptation of plants to unfavorable conditions accounts for the qualitative and quantitative changes of the composition of cell proteins [69,70]. It is well-known that the synthesis of so called stress-proteins dehydrins is activated under stress; they reveal osmolyt type activity and take part in the stabilization of membrane proteins and cell osmotic regulation; thus protecting the cell structures against the oxidative stress [64,70]. Content of total proteins, like proline, was higher in Kvernaqi plants compared to the same species of lagludja ($p \leq 0.03$): in goat's thorn it was 2, 5 times higher, in caper bush 3, 2 times, in spurge 2, 2 times, and in Christ's thorn 1, 6 times higher. Again, the exception was felted germander. Proteins content in this species appeared to be higher in lagludja individuals by 30%, compared to Kvernaqi data (Figure 6).

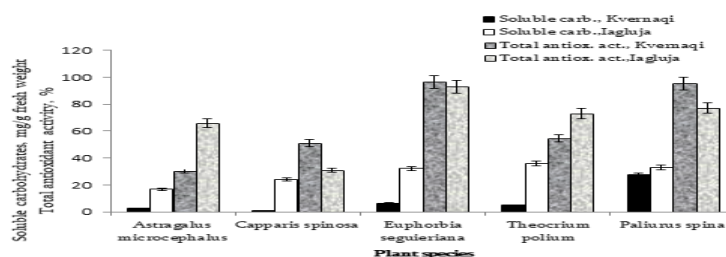
Figure 6. Content of proline and total proteins in leaves of Kvernaqi and Iagluja (East Georgia) plants. Note: (□) Proline, Kvernaqi; (■) Proline, Iagluja; (▨) Proteins, Kvernaqi; (▩) Proteins, Iagluja



It has been established that decrease of the amount of total proteins takes place under various stresses. This is explained by the outflow of nitrogenous substances from the leaf and their synthesis decrease [71]. At the same time the inhibition of photosynthesis and proteins proteolysis may become the reason of total proteins decline [70,72]. Significant decrease of total proteins in Iagluja experimental species may be explained by the decline of nitrates and nitrate reductase activity in their leaves. The increase of total proteins content in felty germander leaves under Iagluja conditions, which we consider to be more extreme than Kvernaqi, may be one of the individual mechanisms of protection against the severe stress. Many authors have mentioned the accumulation of soluble carbohydrates in plant cells in response to various stresses [70,73,74]. These substances decrease the water potential of cells and support the water retention. At the same time they protect proteins from denaturation and account for membrane stability [6,75]. Evidently higher concentration of soluble carbohydrates was established in Iagluja experimental plants, compared to the same species of Kvernaqi hills ($p \leq 0.001$) (Figure 7). The results were 6-times higher in goat's thorn, 20 times higher in caper bush, 5 times in spurge and 1, 2 times higher in Christ's thorn. It is clear that accumulation of such a big amount of soluble carbohydrates under the severe conditions of Iagluja hill is indication to the leading stress-protective function of these substances, while in Kvernaqi hill the protective mechanism of carbohydrates was evident only in Christ's thorn. The total antioxidant activity is a significant characteristic which may be used as a criterion for the evaluation of plant's stress-resistance. The antioxidant activity of the leaves of one and the same plant species appeared to be different by habitats (except the spurge). In goat's thorn and felty germander the Iagluja data prevailed over Kvernaqi ones (2 and 1, 3 times respectively), while in caper bush and Christ's thorn in contrary-the Kvernaqi results were higher (1, 8 and 1, 2 times respectively). According to obtained results the antioxidant activity of caper bush leaves from Iagluja may be regarded as low, that of goat's thorn-moderate, and of spurge, felty germander and Christ's thorn high [76]. Among Kvernaqi species the low antioxidant activity had goat's thorn, moderate-caper bush and felty germander; the high antioxidant activity was revealed in spurge and Christ's thorn. Soil conditions are one of the important factors for normal development of plant. It has been established that the soil microflora significantly accounts for plant's stress-resistance by means of suitable mechanisms, and facilitates their existence under stress conditions [77-80]. Accordingly it may be supposed that local soil conditions play a significant role in

experimental plants' adaptation to stress-conditions of the studied habitats as well. By our opinion, this fact must be taken into account in further observation (Figure 7).

Figure 7. Content of soluble carbohydrates and total antioxidant activity in leaves of Kvernaqi and Iagluja (East Georgia) plants. Note: (■) Soluble carb., Kvernaqi; (□) Soluble carb., Iagluja; (▨) Total antioxid. act., Kvernaqi; (▩) Total antioxid. act., Iagluja



CONCLUSION

In spite of similarity of climatic conditions of Kvernaqi and Iagluja hills, obtained results let assume that Iagluja conditions are more extreme for plants; which may be linked with soil conditions as well. The antioxidant defence mechanisms of experimental species appeared to be partly diverse following habitats. It is evident that stress-adaptive mechanisms have specific peculiarities; although these mechanisms are determined by those factors which plant has to adapt. Activation of the phenol-anthocyanin antioxidant defence mechanism against the stress conditions of both studied habitats was common in all tested plants. Together with the phenol-anthocyanin mechanism of stress-protection, activation of peroxidase was expressed in all tested species of Kvernaqi hill; while in Iagluja accumulation of soluble carbohydrates was evident.

REFERENCES

1. Zhongming Zhu, et al. WMO Provisional Report on the State of the Global Climate. 2021.
2. Allen Craig D, et al. On Underestimation of Global Vulnerability to Tree Mortality and Forest die-off from Hotter Drought in the Anthropocene. Ecosphere. 2015; 8: 1-55.
3. Overpeck J, et al. Dry times ahead. Science. 2010; 328:1642-1643.
4. Garamvolgyi A, et al. Impacts of Climate Change on Vegetation Distribution. Climate Change Induced Vegetation Shifts in the Palearctic Region. Appl Ecol Environ Res. 2013;11: 79-122.

5. Li X and Liu F. Drought Stress Memory and Drought Stress Tolerance in Plants: Biochemical and Molecular Basis. Springer, Cham. 2016;1:17-44.
6. Laxa M Liebthal M, et al. The Role of the Plant Antioxidant System in Drought Tolerance. Antioxidants. 2019;94:1-31.
7. Badridze G, et al. The Biochemical Indices of Drought Resistant Species of Lori Plateau (East Georgia). Int j environ agric res. 2021; 7: 33-49.
8. Kordzakhia M, et al. Climate of Georgia. Ganatleba, Tbilisi. 1971;86.
9. Ukleba D. Encyclopedia. The Main Redaction of Georgian Encyclopedia, Tbilisi. 2018; 662.
10. Amiri MS, et al. Ethnobotanical Knowledge of Astragalus Spp.: The World's Largest Genus of Vascular Plants. Avicenna J Phytomed, 2020;10: 128-142.
11. Li X, et al. A Review of Recent Research Progress on the Astragalus Genus. Molecules, 2014;19:18850-18880.
12. Rafieian Kopaei M, et al. Teucrium polium: Liver and Kidney Effects. J Res Med Sci. 2014;19:478-479.
13. Mahmoudi R, et al. Review on Composition and Antimicrobial Effects of Teucrium (Teucrium polium L.) Grown in Iran and Comparison with the Around the World. J Babol Univ Med Sci. 2017;19:54-56.
14. Candela RG, et al. A Review of the Phytochemistry, Traditional Uses and Biological Activities of the Essential Oils of Genus Teucrium. Planta Med. 2021;87:432-479.
15. USDA. Weed Risk Assessment for Euphorbia falcata L.(Euphorbiaceae) Sickle spurge. 2016.
16. Kemboi D, et al. Review of the Ethnomedicinal Uses, Biological Activities, and Triterpenoids of Euphorbia Species. Molecules, 2020;25:4019.
17. Sakcali MS, et al. Eco-physiology of Capparis spinosa L.: A Plant Suitable for Combating Desertification. Pak J Bot. 2008;40: 1481-1486.
18. Chedraoui S, et al. Capparis Spinosa L. in A Systematic Review: A Xerophilous Species of Multi Values and Promising Potentialities for Agrosystems under the Threat of Global Warming. Front Plant Sci. 2017;8:1845.
19. Brantner A, et al. Antimicrobial activity of Paliurus spina-christi Mill. (Christ's thorn). J Ethnopharmacol. 1996;52:119-122.
20. Zor M, et al. Antigenotoxic Properties of Paliurus Spina-Christi Mill fruits and their Active Compounds. BMC Complement Altern Med. 2017;17:229.
21. Kirichok EI, et al. Ontogenesis of Christ's Thorn (Paliurus spina-christi Mill.). Russ J Ecosyst Ecol. 2018;3:1-21.
22. Ermakov AI. Methods of Plants Biochemical Study. Leningrad, Agropromizdat. 1987;41-42.
23. Pleshkov BP. Practical Handbook in Plant Biochemistry. Moscow, Kolos. 1985; 203-206.
24. Fillipovich IM, et al. Practical Handbook in Biochemistry. Prosveshchenie, Moscow. 1982;40-60.
25. Gavrilenko VF, et al. Big Practical Handbook in Plant Physiology. Visshaia shkola, Moscow. 1975;127-134.

26. Ferraris L, et al. Variations of Phenolics Concentrations as a Consequence of Stress that Induce Resistance to Fusarium wilt of tomato. *J Plant Dis Protect.* 1987;94:624-629.
27. Bates LS, et al. Rapid Determination of Free Proline for Water-stress Studies. *Plant Soil.* 1973;39:205-207.
28. Turkina MV, et al. Methods of Determination of Mono- and Oligosaccharides. In: Pavlinova OA. Ed. *Plant Physiol. Nauka, Moscow.*1971; 20-26.
29. Danilova NS. Determination of Nitrates in Plant Material. *Plant physiology.* 1963;10:497-498.
30. Koleva II, et al. Screening of Plant Extracts for Antioxidant Activity: A comparative Study on Three Testing Methods. *Phytochem Anal.* 2002;13:8-17.
31. Ananthi K, et al. Soluble protein, Nitrate Reductase, and Yield Response in Cotton Genotypes under Water Stress. *Insight Biochemistry.* 2012; 2:1-4.
32. Mc Carthy J K, et al. Nitrate Reductase Knockout Uncouples Nitrate Transport from Nitrate Assimilation and Drives Repartitioning of Carbon Flux in a Model Pennate Diatom. *Plant Cell.* 2017;29:2047–2070.
33. Sinay H, et al. Nitrate Reductase Activity of Seven Local Corn Cultivars from South West Maluku District During Water Stress Caused by Polyethylene Glycol 6000 under Green House Condition. *Int J Biosci.* 2019;15:80-86.
34. Kapoor D, et al. The Impact of Drought in Plant Metabolism: How to Exploit Tolerance Mechanisms to Increase Crop Production. *Appl Sci.* 2020;10, 5692.
35. Nicodemus MA, et al. Nitrate Reductase Activity and Nitrogen Compounds in Xylem Exudate of *Juglans Nigra* Seedlings: Relation to Nitrogen Source and Supply. *Trees,* 2008;22:685-695.
36. Dias DN. Patterns of Nitrate Reductase Activity Vary According to the Plantfunctional Group in a Mediterranean Maquis. *Plant Soil,* 2011;347:363-376.
37. Correia MJ, et al. Effects of Water Deficit on the Activity of Nitrate Reductase and Content of Sugars, Nitrate and Free amino Acids in the Leaves and Roots of Sunflower and White Lupin Plants Growing under Two Nutrient Supply Regimes. *Physiologia Plantarum,* 2005;124:61-70.
38. Ommen OE,et al. Chlorophyll Content of Spring Wheat Flag Leaves Grown under Elevated CO₂ Concentrations and Other Environmental Stresses within the ESPACE-wheat project. *Eur J Agron:* 1999; 10:197-203.
39. Herbinger K, et al. Complex Interactive Effects of Drought and Ozone Stress on the Antioxidant Defence Systems of two Wheat Cultivars. *Plant Physiol Biochem.* 2002; 40: 691-696.
40. Ma Y, et al. Drought and Salinity Stress Responses and Microbe-Induced Tolerance in Plants. *Front Plant Sci.* 2020;11:591911.
41. Chupakhina GN, et al. Natural Antioxidants (ecological aspect). *Baltic Federal university, Kaliningrad.* 2011; 111.
42. Noctor G, et al. The Roles of Reactive Oxygen Metabolism in Drought: Not So Cut and Dried. *Plant Physiology,* 2014;164:1636-1648.
43. Lopez-Huertas E, et al. Stress Induces Peroxisome Biogenesis Genes. *EMBO J.* 2000; 19: 6770-6777.

44. Harinasut P, et al. Salt Effects on Antioxidant Enzymes in Mulberry Cultivar. *Science Asia*, 2003;29:109-113.
45. Kolupaev IE, et al. Antioxidant System and Plant Resistance to Water Deficiency. *Plant Physiology*, 2019;51:28-54.
46. Mhamdi A, et al. Catalase Function in Plants: A focus on Arabidopsis Mutants as Stress-Mimic Models. *J Exp Bot*. 2010; 61:4197-4220.
47. Chakrabarty A, et al. Antioxidant signaling and Redox Regulation in Drought and Salinity-Stressed Plants. *Afr J Plant Sci*. 2016;1:465-498.
48. Venkatesh J, et al. Role of L-ascorbate in Alleviating Abiotic Stresses in Crop Plants. *Bot Studies*, 2014;38:55.
49. Yang Y, et al. Effect of Drought and Low Light on Growth and Enzymatic antioxidant System of *Picea Asperata* Seedlings. *Acta Physiol Plant*. 2008;30: 433-440.
50. Singh S, et al. Differential Responses of Antioxidative Defence System to Long-Term Field Drought in Wheat (*Triticum aestivum* L.) Genotypes Differing in Drought Tolerance. *J Agron Crop Sci*. 2012;198:185-195.
51. Abbasi AR, et al. Specific Roles of Alpha- and Gamma-Tocopherol in Abiotic Stress Responses of Transgenictobacco. *Plant Physiol*. 2007;143: 1720-1738.
52. Giacomelli L, et al. Arabidopsis *Thaliana* Deficient in two Chloroplast Ascorbate Peroxidases Shows Accelerated Light-Induced Necrosis when Levels of Cellular Ascorbate are Low. *Plant Mol Biol*. 2007;65: 627-644.
53. Mullineaux PM, et al. Spatial Dependence for Hydrogen Peroxide-Directed Signaling in Light-Stressed Plants. *Plant Physiol*. 2006;141: 346-350.
54. Gill SS, et al. Reactive Oxygen Species and Antioxidant Machinery in Abiotic Stress Tolerance in Crop Plants. *Plant Physiol Biochem*. 2010; 48: 909-930.
55. Winkel-Shirley B. Biosynthesis of Flavonoids and Effects of Stress. *Curr Opin Plant Biol*. 2002;5:218-223.
56. Cesar G, et al. *Plant Phenolics and Human Health: Biochemistry, Nutrition, and Pharmacology*, John Wiley and Sons, Inc. 2010;1-116.
57. Sharma A, et al. Response of Phenylpropanoid Pathway and the Role of Polyphenols in Plants under Abiotic Stress. *Molecules*. 2019;24:2452.
58. Gould KS, et al. When are Foliar Anthocyanins Useful to Plants? Re-evaluation of the Photoprotection Hypothesis using Arabidopsis *Thaliana* Mutants that Differ in Anthocyanin Accumulation. *Environ Exp Bot*. 2018;154:11-22.
59. Kovinich N, et al. Abiotic stresses Induce Different Localizations of Anthocyanins in Arabidopsis. *Plant Signal Behav*. 2015;10:7.
60. Kamjad Y, et al. Effects of Drought Stress on Anthocyanin Accumulation in Mulberry Fruits. *Asian J Plant Sci*. 2021;20: 450-460.

61. Chalker-Scott L. Do Anthocyanins Function as Osmoregulators in Leaf tissues? Academic Press. 2002;37:103-127.
62. Szabados L, et al. Proline: A Multifunctional Amino Acid. Trends Plant Sci, 2009; 15: 89-97.
63. Meng J, et al. The Ameliorative Effects of Exogenous Melatonin on Grape Cuttings under Water-Deficient Stress: Antioxidant Metabolites, Leaf Anatomy, and Chloroplast Morphology. J Pineal Res. 2014;57:200-212.
64. Iqbal A, et al. High Nitrogen Enhance Drought Tolerance in Cotton through Antioxidant Enzymatic Activities, Nitrogen Metabolism and Osmotic Adjustment. Plants. 2020;9:178.
65. Kaur G and Asthir B.. Proline: A key Player in Plant Abiotic Stress Tolerance. Biol Plant. 2015;59:609-619.
66. Ashraf MA, et al. Recent Advances in Abiotic Stress Tolerance of Plants Through Chemical Priming: An Overview. seed priming. 2018;51-79.
67. Kartashov AV. Significance of Morpho-Physiological Peculiarities of Plantago Major L. and P. Maritima L. in Supporting of Water-Salt Balance under Salinization. A thesis. 2013;23.
68. Joseph EA, Radhakrishnan VV and Mohanan KV. A Study on the Accumulation of Proline an Osmoprotectant Amino Acid under Salt Stress in Some Native Rice Cultivars of North Kerala, India. Univ J Agr Res. 2015;3:15-22.
69. Parida A K, et al. Alterations in Photosynthetic Pigments, Protein and Osmotic Components in Cotton Genotypes Subjected to Short-term Drought Stress Followed by Recovery. Plant Biotechnol Rep. 2007;1:37-48.
70. Mohammadkhani N, et al. Effects of Drought Stress on Soluble Proteins in two Maize Varieties. Turk J Biol. 2008;32:23-30.
71. Sorkheh K, et al. Salt Stress Induction of Some Key Antioxidant Enzymes and Metabolites in Eight Iranian wild Almondspecies. Acta Physiol Plant. 2012;34:203-213.
72. Taiz L, et al. Plant Physiology and Development. Sixth Edition published by Sinauer Associates, Sunderland, USA. 2016;761.
73. Prado FE, et al. Effect of NaCl on Germination, Growth and Soluble Sugar Content in Chenopodium quinoa Willd Seeds. Bot. Bull. Acad. Sin. 2000; 41: 27-34.
74. Finkelstein RR, et al. ABA and Sugar Interactions Regulating Development: Cross-talk or Voices in a Crowd. Curr Opin Plant Biol. 20015;1:26-32.
75. Couee I, et al. Involvement of Soluble Sugars in Reactive Oxygen Species Balance and Responses to Oxidative Stress in Plants. J Exp Bot. 2006;57:449-459.
76. Luzia D M, et al. Study of Antioxidant Activity of Non-Conventional Brazilian fruits. J Food Sci Techno. 2014; 51:1167-1172.
77. Bardi L, et al. Drought and Nutritional Stresses in Plant: Alleviating Role of Rhizospheric Microorganisms. In: abiotic stresses . 2012;1-57.
78. Hossain A, et al. The Role of Soil Microorganisms in Plant Adaptation to Abiotic Stresses: Current Scenario and future perspectives. Glob Chang Biol. 2022;233-278.

79. Lowry OH, et al. Protein Measurement with the Folin Phenol Reagent. *J Biol Chem.* 1951;139:256-275.
80. Mc Dowell N, et al. Mechanisms of Plant Survival and Mortality During Drought: Why do Some Plants Survive while Others Succumb to Drought? Tansley review. *New Phytologist.* 2008;178:719-739.