Future Resources: Microalgal Biotechnology

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Review Article

ABSTRACT

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Keywords: Microalgae, Photobioreactors, Nutrients New innovative advancements consider a noteworthy reduction in the assets fundamental for microalgae creation. This could prompt an expansion in the utilization of microalgae in an extensive variety of uses - from sustenance creation to medical applications and nutrient recovery.

INTRODUCTION

Numerous years of exploration work on the improvement of photobioreactors, which use photosynthesis to transform light vitality into biomass, have gone before this achievement. The supposed "Permeable Substrate Bioreactor" (PSBR), otherwise called the twin-layer framework, utilizes another standard to isolate the green growth from a supplement arrangement by method for a permeable reactor surface on which the microalgae are caught in biofilms ^[1-10]. Unique about this new methodology is that it lessens the measure of fluid required in contrast with the presently utilized innovation, which develops green growth in suspensions, by a component of up to one hundred. The PSBR technique along these lines considers a noteworthy diminishment in vitality and for an expansion in the arrangement of green growth that can be developed ^[11-20].

Current triumphs in PSBR advancement and the ascent in enthusiasm for this innovation as of late could flag a turn in the origination of future photobioreactors in microalgae biotechnology.

Microalgae have various applications: they are customary wellsprings of protein and starches. They can likewise be utilized as a part of the maintainable generation of regular colors and cell reinforcements, for example, beta-carotene and astaxanthin. Polyunsaturated unsaturated fats, which normally originate from fish oil, can likewise be integrated from microalgae ^[21-30]. Besides, green growth can serve as the premise for pharmaceutical operators, for example, antiviral and anticancerogenous substances. In natural biotechnology, new ideas are at present being created to utilize microalgae to recoup phosphor and nitrogen from sewage and reintroduce them into the supplement cycle by method for natural manures ^[31-40].

There are no less than 30,000 known types of microalgae. Just a modest bunch are presently of business importance. These are developed for extraction of high-esteem parts, for example, shades or proteins. A couple of animal groups are utilized for nourishing shellfish or other aquaculture purposes. One of the key examination undertakings for commercialization of green growth for vitality reasons for existing is to screen species for good creation and for simplicity of development and preparing, among other criteria. The primary center of screening is as of now on lipid efficiency, and resulting esterification, however maturation choices ought not be overlooked. Most screening projects incorporate freshwater species. There is no agreement concerning ideal frameworks for

microalgae development. Researchers differ about whether open or shut or some blend of development frameworks is generally good. Open-lake frameworks, such as raceways, entail low capital and operating cost, but also low productivity and lack of control over development. Shut frameworks, for example, photograph bioreactors (PBR) are significantly costlier however offer higher profitability. In existing business applications, simulated light and in some cases warmth are utilized. This can be advocated on a little scale for high-esteem item make. For vitality purposes, just normal light and once in a while waste warmth ought to be considered. The greatest obscure in Ireland or other comparable atmospheres is whether it is conceivable to accomplish sensible profitability in perspective of winning regular light and temperatures. For areas at higher scope, it might be conceivable to recognize neighborhood strains requiring low light powers and lower water temperatures however giving acceptable development rates and yields ^[41-50].

Fleeting development rate is regularly erroneously extrapolated to yearly profitability. It is likely that a substantial regularity punishment will exist if microalgae are to be developed in Ireland where the scope is 53°N. Inspite of this confinement, microalgae generation for biofuel can't be discounted without further research and approval of the idea in Ireland. Partners in Ireland from the scholarly, mechanical and entrepreneurial group wish to show this innovation. An idealistic situation is sketched out inside the report where 100 ha of microalgae generation is accomplished by 2020. A few critical examination advances would be required if this somehow managed to be accomplished ^[51-60].

There is an agreement that the photosynthetic effectiveness of earthbound plants is 1% or less. Promoters of green growth innovation for biofuel expect the breaking points of microalgae photosynthetic proficiency to be pushed out to some place somewhere around 3 and 6%. 6% can be set as a flat out most extreme hypothetical effectiveness that is unrealistic to ever be acquired under genuine conditions. Efficiency claims for microalgae frameworks are frequently exaggerated ^[61-65].

Considering the essentials of photosynthesis, anything above 53 t/ha/yr of dry biomass in the Irish atmosphere ought to be treated with alert. An examination objective for Ireland could be to exhibit biomass areal efficiency rates of 25 dry t/ha/yr and to acquire 25% of valuable lipids, yielding 6.25 m3/ha/yr. Desires ought to be humble until at any rate these preparatory targets are met ^[66-70].

Supplements and carbon are other key necessities for microalgal development. For carbon, fumes gas from force plants which contain critical amounts of minimal effort CO_2 can be utilized. This is a piece of the plan of action of most biofuel ventures, which additionally permits power plants to reuse CO_2 . Algal slurry is 15-25% dry weight after accumulation. Dry lipids are important for esterification and evacuation of water is costly. Advancement of lipase for direct esterification or other extraction strategies could evacuate the drying step. Unsaturated fat substance is high in algal oils and their nearness brings down esterification yields ^[71-80].

Microalgae have huge lipid content and even high lipid content under certain anxiety conditions. Research labs have demonstrated that some microalgae strains can generate 70% lipid in their biomass. Be that as it may this has not yet been found in genuine conditions where most extreme yields of 30% are experienced. There might be open doors for applying iorefinery-sort procedures to concentrate and separate a few business items from microalgal biomass. Other than lipids, microalgal biomass offers open doors for getting extra business materials. These incorporate maturations to get ethanol and biogas. It is likewise conceivable to deliver protein-rich food for both creature and human utilization ^[81-90].

Poly unsaturated fats (PUFAs) are a potential co-result of biodiesel generation from microalgae. PUFAs are a vegetable source contrasting option to e.g. fish oils and different oils rich in omega-3 unsaturated fats. Mass markets for the co-items conceivably accessible by means of a biorefinery procedure have not been illustrated, and this is an examination need if the biorefinery idea is to demonstrate a legitimate business model. Due to the little beginning size of any pilot creation of microalgal oil, it is likely that feedstocks would first be utilized as a part of existing biodiesel refineries keeping in mind the end goal to trial the idea. The flying business is especially intrigued by algal biodiesel, because of its prevalent frosty temperature execution, vitality thickness and capacity dependability. Current development costs just legitimize extraction of high-esteem specialty parts. A diminishment by no less than an element of five is important to make microalgae appealing for their lipid content ^[91-95].

There is noteworthy action around the world, with news about ventures and research programs rising on a consistent schedule. There are liable to be more than 30 new US patent applications submitted amid 2008, which will surpass the aggregate for the former 6 years. The low number of licenses is favorable position for specialists and potential speculators as it leaves open doors for further advancements and development security.

Ebb and flow Irish examination exercises stay unassuming in the global setting. Unless the key test of acquiring microalgae suited to the Irish atmosphere is fathomed, this is liable to remain the case. About 79 TJ could originate from microalgae assets by 2020 considering the most idealistic situation produced for the reasons for this report. This is a small amount of 1% of national street fuel request ^{[96-99].}

CONCLUSION

The vitality commitment from marine green growth by 2020 is prone to be unobtrusive. The open doors for era of innovation, turn off movement, occupations, venture and the potential for new licensed innovation creation have not been considered in this audit. The enthusiasm for non-vitality items, for example, nutraceuticals, shades, proteins, useful sustenance's and other synthetic constituents is right now financially more critical than vitality.

REFERENCES

- 1. Beijerinck MW. Culturversuche mit Zoochlorellen, Lichenengonidien und anderen niederen Algen. Bot.Zeitung (1890) 48: 725–739.
- 2. Warburg O. Über die Geschwindigkeit der photochemischen Kohlensäurezersetzung in lebenden Zellen. Biochem. Z (1919) 100: 230–270.
- 3. Seckbach J and Oren A Microbial Mats-Modern and Ancient Microorganisms in Stratified Systems. Springer, 2010.
- Jhala YK, et al. Biodiversity of Endorhizospheric Plant Growth Promoting Bacteria. J Biofertil Biopestici. 2015; 6:151.
- 5. Du RY, et al. Effect of Bacterial Application on Metal Availability and Plant Growth in Farmland-Contaminated Soils. J Bioremed Biodeg. 2016; 7:341.
- 6. Abdeljalil NOB, et al. Bio-suppression of Sclerotinia Stem Rot of Tomato and Biostimulation of Plant Growth
- 7. Using Tomato-associated Rhizobacteria. J Plant Pathol Microbiol. 2016; 7:331
- 8. Varshney S, et al. Contribution of Plant Growth Regulators in Mitigation of Herbicidal Stress. J Plant Biochem Physiol. 2015; 3:160.
- 9. Wan J, et al. Potential Application of Chitin Signaling in Engineering Broad-Spectrum Disease Resistance to Fungal and Bacterial Pathogens in Plants. Adv Crop Sci Tech. 2013;1: e103.
- 10. Chauhan A. Sinorhizobium melilotiBacteria Contributing to Rehabilitate the Toxic Environment. J Bioremed Biodeg. 2015;6: e164.
- 11. El-Halmouch Y et al. The Potential of Cell-free Cultures of Rhizobium leguminosarum, Azotobacter chroococcumand Compost Tea as Biocontrol Agents for Faba Bean Broomrape Orobanche crenata Forsk. J Plant Pathol Microb. 2013; 4:205.
- 12. Abdeljalil NOB, et al. Characterization of Tomato-associated Rhizobacteria Recovered from Various Tomatogrowing Sites in Tunisia. J Plant Pathol Microbiol. 2016; 7:351.
- 13. Okon E, et al. Evaluation and Characterisation of Composite Mesoporous Membrane for Lactic Acid and Ethanol Esterification. J Adv Chem Eng. 2016; 6:147.
- 14. Sanalibaba Pand Çakmak GA. Exopolysaccharides Production by Lactic Acid Bacteria. Appli Micro Open Access. 2016; 2:115.
- 15. Chatterjee M, et al. Effect of Fruit Pectin on Growth of Lactic Acid Bacteria. J Prob Health. 2016; 4:147.
- 16. Liu YH et al. Inhibitory Effect of Lactic Acid Bacteria on Uropathogenic Escherichia Coli-Induced Urinary Tract Infections. J Prob Health. 2016; 4:144.
- 17. Anacarso I, et al. Amoebicidal Effects of Three Bacteriocin like Substances from Lactic Acid Bacteria against Acanthamoeba polyphaga. J Bacteriol Parasitol. 2014; 5:201.
- 18. Limanska N, et al. Study of the Potential Application of Lactic Acid Bacteria in the Control of Infection Caused by Agrobacterium tumefaciens. J Plant Pathol Microb. 2015; 6:292.
- 19. Wedajo B. Lactic Acid Bacteria: Benefits. Selection Criteria and Probiotic Potential in Fermented Food. J Prob Health. 2015; 3:129.

Research and Reviews: Journal of Chemistry

- 20. Singh H, et al. Lactic acid Bacteria Isolated from Raw Milk Cheeses: Ribotyping. Antimicrobial Activity against Selected Food Pathogens and Resistance to Common Antibiotics. J Food Process Technol. 2015; 6:485.
- 21. Miranda JM, et al. Technological Characterization of Lactic Acid Bacteria Isolated from Beef Stored on Vacuum-Packaged and Advanced Vacuum Skin Packaged System. J Food ProcessTechnol. 2014; 5:338.
- 22. Sivaramasamy E, et al. Enhancement of Vibriosis Resistance in Litopenaeus vannameiby Supplementation of Biomastered Silver Nanoparticles by Bacillus subtilis. J Nanomed Nanotechnol. 2016; 7:352.
- 23. Kalarani V, et al. Effect of Dietary Supplementation of Bacillus subtilisand Terribacillus saccharophilluson Innate Immune Responses of a Tropical Freshwater Fish,Labeo rohita. J Clin Cell Immunol. 2016; 7:395.
- 24. Javed S, et al. Hyper-production of Alkaline Protease by Mutagenic Treatment of Bacillus subtilisM-9 using Agroindustrial Wastes in Submerged Fermentation. J Microb Biochem Technol. 2013; 5:074-080.
- 25. Ping SP, et al. Effect of Isoflavone Aglycone Content and Antioxidation Activity in Natto by Various Cultures of Bacillus SubtilisDuring the Fermentation Period. 2012.
- 26. Tam NKM, et al. The Intestinal Life Cycle of Bacillus subtilis and Close Relatives. J Bacteriol. 2006; 188:2692-2700.
- 27. Anochie PI, et al. Tuberculosis and Human Immunodeficiency Virus Co-Infection in Rural Eastern Nigeria. J Med Diagn Meth 2013; 2:118.
- 28. Soula F, et al. Physical Activity Participation and Cardiovascular Fitness in People Living with Human Immunodeficiency Virus: A One-Year Longitudinal Study. J AIDS Clinic Res 2013; S9:002.
- 29. Atrio J, et al. Contraceptive use in Adolescents with Perinatally and Behaviorally Acquired Human Immunodeficiency Virus Infection Seen in a Public Los Angeles County Clinic. J AIDS Clin Res2013;4: 244.
- 30. Krol M, et al. Increased Exhaled Hydrogen Peroxide in Human Immunodeficiency Virus-Infected Patients without
- 31. Clinical Signs and Symptoms of Opportunistic Lung Disease. J AIDS Clinic Res2012; 3:183.
- 32. Bismara BA, et al. Antiretroviral Drug Resistance in Brazilian Children Infected by Human Immunodeficiency Virus Type 1. J Antivir Antiretrovir 2012; 4: 066-074.
- 33. Neurath AR, et al.Prevention of Human Immunodeficiency Virus Type 1 Transmission by Pharmaceuticals Targeted to Host Proteins Required for Virus Infection? Consideration of Farnesyl Thiosalicylic Acid, a Ras Inhibitor. J Antivir Antiretrovir 2009; 1: 072-075.
- 34. Pavlova-McCalla E, et al. Socioeconomic Status and Survival of People with Human Immunodeficiency Virus Infection before and after the Introduction of Highly Active Antiretroviral Therapy: A Systematic Literature Review. J AIDS Clinic Res 2012; 3:163.
- 35. Lifson AR, et al. Use of Community Health Support Workers for Persons Living with Human Immunodeficiency Virus in RuralEthiopia: Lessons Learned. J AIDS Clin Res 2014; 5:324.
- 36. Patekar D, et al.Prevalence of Viral Coinfections with EBV and CMV and Its Correlation with CD4 Count In HIV-1 Serpositive Patients. J AIDS Clin Res2015; 6:520.
- 37. Sangarei, et al. CoinfectionHIV and Malaria in Department of Paediatrics of the University Hospital Souro Sanou. J Hematol Thrombo Dis 2015; 3:213.
- 38. Lenjisa JL, et al. Mortality of Adults on Antiretroviral Therapy with and without TB co-infection in Jimma University Hospital, Ethiopia: Retrospective Cohort Study. J AIDS Clin Res 2014; 5:350.
- 39. de Azevedo RG, et al. HIV/HCV Coinfection: Considerations about Treatment. J Med Microb Diagn 2014; 3:148.
- 40. Toru Shizuma Efficacy of Treatments for Patients with Chronic Liver Disease due to Coinfection with Hepatitis B and C Viruses. J Gastroint Dig Syst 2014; 4:181.
- 41. Tobias C, et al. Makingthe connection: the importance of engagement and retention in HIV medical care. AIDS Patient Care STDS 21 Suppl2007;1: S3–S8.
- 42. Moanna A, et al. Primary Human Immunodeficiency Virus Infection and Rhabdomyolysis. J AIDS Clinic Res 2011; 2:119.
- 43. Montesano C, et al. Impact of Human Leukocyte Antigen Polymorphisms in Human Immunodeficiency Virus Progression in a Paediatric Cohort Infected with a Mono-phyletic Human Immunodeficiency Virus-1 Strain. J AIDS Clin Res2014; 5:282.
- 44. Ibeh IN, et al. Evaluation of the Anti-Human Immunodeficiency Virus Hiv Properties of Dxl Decoction X-Liquid-Bioclean Ii. J Clinic Toxicol 2013; S12:004.

Research and Reviews: Journal of Chemistry

- 45. Aleme H, et al. Sereoprevalence of Immunoglobulin-G and of Immunoglobulin-M Anti-Toxoplasma gondiiAntibodies in Human Immunodeficiency Virus Infection/Acquired Immunodeficiency Syndrome Patients at Tikur Anbessa Specialized Hospital, Addis Ababa, Ethiopia. J Infect Dis Ther 2013; 1:119.
- 46. Adams J, et al. Plasma Cell Myeloma with Unusual Expression of CD19, CD10, CD45 and Surface Light Chain in a Human Immunodeficiency Virus Positive Patient. J Leuk Los Angel 2013;1: 126.
- 47. Grulich AE, et al. Incidence of cancers in people with HIV/AIDS compared with immunosuppressed transplant recipients: a meta-analysis. Lancet2007; 370:59–67.
- 48. Engels EA, et al. Cancer risk in people infected with human immunodeficiency virus in the United States.InternationalJournalofCancer2008;123:187–194.
- 49. Silverberg MJ, et al. HIV infection and the risk of cancers with and without a known infectious cause. AIDS 2009; 23:2337–2345.
- 50. AngelettiPC, et al. The viral etiology of AIDS-associated malignancies. AdvancesinPharmacology2008; 56:509– 557.
- 51. Engels EA, et al. Trends in cancer risk among people with AIDS in the United States 1980-2002.AIDS2006; 20:1645-1654.
- 52. Carbone A, et al. HIV-associated Hodgkin lymphoma. Curr Opin HIV AIDS 2009; 4:3–10.
- 53. Silverberg MJ, et al. AIDS-defining and non-AIDS-defining malignancies: cancer occurrence in the antiretroviral therapy era. CurrentOpinioninOncology2007; 19:446–451.
- 54. GroggKL, et al. HIV infection and lymphoma. JClinPathol2007; 60:1365-1372.
- 55. Simard EP, et al. Spectrum of cancer risk late after AIDS onset in the United States.ArchInternMed2010; 170 :1337–1345.Shiels MS, et al. Age at cancer diagnosis among persons with AIDS in the United States.AnnInternMed2010; 153:452–460.
- 56. Spano JP, et al. AIDS-related malignancies: state of the art and therapeutic challenges. Journal of Clinical Oncology 2008;26: 4834–4842.
- 57. Heard I Prevention of cervical cancerin women with HIV.Current Opinion in HIV and AIDS2009; 4: 68–73.
- 58. Nguyen VM, et al. How Long Is Too Long in Contemporary Peer Review? Perspectives from Authors Publishing in Conservation Biology Journals. PLoS One. 2015; 10: e0132557.
- 59. Salasche SJ. How to "peer review" a medical journal manuscript. Dermatol Surg. 1997; 23: 423-428.
- 60. Roberts J. An Author's Guide to Publication Ethics: A Review of Emerging Standards in Biomedical Journals. Headache: The Journal of Head and Face Pain. 2009; 49: 578-589.
- 61. Kempers RD. Ethical issues in biomedical publications. Fertil Steril. 2002; 77: 883-888.
- 62. Reyes H. Honesty and good faith: Two cornerstones in the ethics of biomedical Publications. Rev Méd Chile. 2007; 135: 415-418.
- 63. Campanario JM. El sistema de revision por expertos (peer-review): muchos problemas y pocas soluciones. Revista española de Documentación Científica. 2002; 25: 267-285.
- 64. Zhu J, et al. Evaluating the Pros and Cons of Different Peer Review Policies via Simulation. Sci Eng Ethics. 2015.
- 65. Shrivastava S, Shrivastav A, Sharma J (2016) Co-exposure Effects of Selenium and Mercury on Phaseolus vulgaris Excised Leaves Segment by Enhancing the NR, Anti-oxidative Enzyme Activity and Detoxification Mechanisms Adv Tech Biol Med 4:178.
- 66. Yanagimachi R, et al. The use of zona-free animal ova as a test-system for the assessment of the fertilizing capacity of human spermatozoa. Biol Reprod 1976; 15: 471-476.
- 67. Huang JM, et al. Studies on the integration of hepatitis B virus DNA sequence in human sperm chromosomes. Asian J Androl 2002; 4: 209-212.
- 68. Ali BA, et al. Expression of hepatitis B virus genes in early embryonic cells originated from hamster ova and human spermatozoa transfected with the complete viral genome. Asian J Androl 2006; 8: 273-279.
- 69. Lee WM. Hepatitis B virus infection. N Engl J Med 1997; 337: 1733-1745.
- 70. Ahmed MM, et al. An improved experimental model for studying vertical transmission of hepatitis B virus via human spermatozoa. J Virol Methods 2008; 151: 116-120.
- 71. Sambrook J, et al. Molecular cloning. A laboratory manual. 3rd ed. New york: Clod Spring Habor Laboratory Press 2001; 1:21-1.83.
- 72. Davison F, et al. (1987) Detection of hepatitis B virus DNA in spermatozoa, urine, saliva and leucocytes, of chronic HBsAg carriers. A lack of relationship with serum markers of replication. J Hepatol 4: 37-44.

Research and Reviews: Journal of Chemistry

- 73. Lang ZW. Distribution of hepatitis B virus in testicle tissue in patients with hepatitis B infection. Zhonghua Yi Xue Za Zhi 1993; 73: 329-331.
- 74. Ganem D and Prince AM. Hepatitis B virus infection-natural history and clinical consequences. N Engl J Med 2004; 350: 1118-1129.
- 75. Mann S, et al. Sol-Gel Synthesis of Organized Matter. Chem Mater. 1997; 9:2300-2310.
- 76. Datta KKR, et al. Aminoclay: a functional layered material with multifaceted applications. J Mater Chem. 2013; A1:6707-6718.
- 77. Choi MH, et al. Aquatic ecotoxicity effect of engineered aminoclay nanoparticles. Ecotox Environ. Safe. 2014; 102:34-41.
- 78. Lee YC, et al. Utilizing the algicidal activity of aminoclay as a practical treatment for toxic red tides. Sci Rep. 2013; 3:1292.
- 79. Yang L, et al. Biodistribution and clearance of aminoclay nanoparticles: implication for in vivo applicability as a tailor-made drug delivery carrier. J Mater Chem. 2014; 2:7567-7574.
- 80. Lee YC, et al. Optical properties of fluorescein-labeled organoclay. Photochem Photobiol. 2010; 86:520-527.
- 81. Ji HM, et al. Efficient harvesting of wet blue-green microalgal biomass by two-aminoclay [AC]-mixture systems. Bioresour Technol. 2016; 211:313-318.
- 82. Chaturbedy P, et al. pH-Sensitive breathing of clay within the polyelectrolyte matrix. ACS Nano 2010; 4:5921-5929.
- 83. Rhim JW, et al. Bio-nanocomposites for food packaging applications. Prog Polym Sci. 2013; 38:1629-1652.
- 84. Chandrasekaran G, et al. Antimicrobial activity of delaminated aminopropyl functionalized magnesium phyllosilicates. Appl Clay Sci. 2011; 53:729-736.
- 85. Lee YC, et al. Dual-end functionalized magnesium organo-(phyllo)silicates via co-condensation and its antimicrobial activity. Appl Clay Sci. 2013; 84:474-485.
- 86. Jung SW, et al. Can the algicidal material Ca-aminoclay be harmful when applied to a natural ecosystem? An assessment using microcosms. J Hazard Mater. 2015; 298:178-187.
- 87. Farooq W, et al. Efficient microalgae harvesting by organo-building blocks of nanoclays. Green Chem. 2013; 15:749-755.
- 88. Lee YC, et al. Harvesting of oleaginous Chlorella sp. by organoclays. Bioresour Technol. 2013; 132:440-445.
- 89. Kim S, et al. A Simple and Non-Invasive Method for Nuclear Transformation of Intact-walled Chlamydomonas reinhardtii. PLoS One. 2014;9: e101-118.
- 90. Kang KS, et al. In-vitro cytotoxicity assessment of carbon-nanodot-conjugated Fe-aminoclay (CD-FeAC) and its bio-imaging applications. J Nanobiotechnology. 2015; 13:88.
- 91. Lee YC, et al. Oil extraction by aminoparticle-based H2O2 activation via wet microalgae harvesting. RSC Adv. 2013; 3:12802-12809.
- 92. Han HK, et al. Magnesium and Calcium Organophyllosilicates: Synthesis and In vitro Cytotoxicity Study. ACS Appl. Mater. Interfaces. 2011; 3:2564-2572.
- 93. Christaki E, Bonos E, Florou-Paneri P (2015) Innovative microalgae pigments as functional ingredients in nutrition. In: Kim SK (ed.) Handbook of Marine Microalgae: Biotechnology Advances. Elsevier Academic Press, London, UK, pp: 233-243.
- 94. Mulders KJM, Lamers PP, Martens DE, Wijffels RH (2014) Phototropic pigment production with microalgae: Biological constraints and opportunities. J Phycol 50: 229-242.
- 95. Juan Bueno. Antimicrobial Adjuvants Drug Discovery, the Challenge of Avoid the Resistance and Recover the Susceptibility of Multidrug-Resistant Strains. J Microb Biochem Technol 2016; 8: 169-176.
- 96. Titanji VPK and Assam JPA. Drug Sensitivity and Molecular Diversity of M. tuberculosis in Cameroon: A Metaanalysis. J Vasc Med Surg 2016; 4: 266.
- 97. Amin A. Al-Sulami, et al. Frequency of Rapid Growing Mycobacteria among Tuberculosis Suspected Patients in Basra-Iraq. Biol Med (Aligarh) 2016;8: 297.
- 98. Jena L, et al. Computational Approach in Understanding Mechanism of Action of Isoniazid and Drug Resistance. Mycobact Dis 2016; 6: 202.
- 99. Parameswaran S and Sanjukta P. Rv3802c in Tuberculosis Therapeutics. Mycobact Dis 2016; 6: 204.